Diversity and distribution of ciliated protozoans on the mangrove leaf litters of *Rhizophora apiculata*

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Abstract



The diversity and distribution of ciliated protozoans on the mangrove leaf litters of *Rhizophora apiculata* with physicochemical parameters were studied for one year (June 2014 - May 2015) from ten selected stations of the Ayiramthengu mangrove ecosystem in Kerala, India. Taxonomic studies recorded 115 species of ciliates, belonging to 16 subclasses among them Peritricha represents the subclass having the maximum species number and abundance followed by Suctoria. *Vorticella companula* Ehrenberg, 1831 records the maximum abundance and dominance index followed by *Euplotes minuta*. Multivariate statistical analysis and analysis of variance revealed seasonal difference in the distribution of mangrove ciliates, where the highest number of species were distributed in post-monsoon and minimum in premonsoon. Canonical correspondence analysis explains the importance of dissolved oxygen, temperature, conductivity, salinity, sulphate, phosphate and nitrate in the distribution and abundance of ciliates protozoans. The study highlights the seasonal difference in the distribution, diversity and abundance of ciliated protozoans on the mangrove leaf litters of *Rhizophora apiculata* in the Ayiramthengu mangrove ecosystem, Kerala, India.

Keywords: Ciliates, Leaf litter, Peritricha, Spirotrichea

Introduction

Mangroves are a group of salt tolerant flowering trees and shrubs, which are circumglobal in distribution and are specially adapted to grow in the intertidal regions of the tropical and subtropical coastlines (Tomlinson 1986; Duke 1993; Hogarth 1999; Saenger 2002). Five per cent of world's mangrove vegetation is distributed in India, spreading over 4,500 km² along the coastal states. Earlier, Kerala was abundant with 700 km² of mangrove forest and the deterioration started in the middle of the 20th century. At present, the mangrove vegetation has been reduced to a mere 17 km² that comes under the control of state government (Mini *et al.* 2014).

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Total of 15 pure and 33 semi mangrove species had been recorded from entire costal area of Kerala (Vidyasagaran et al. 2014). Mangrove flora of Ayiramthengu comprise of totally nine species belonging to six families (Myrsinaceae, Avicenniaceae, Rhizophoraceae. Euphorbiacea. Combretaceae and Sonneratiaceae), Avicenniaceae is the largest family in Ayiramthengu region followed by Rhizophoraceae (Vijayan et al. 2015). The amount of leaf litter produced by mangrove plants will not be same as in every species, it will vary with leaf size, wide and dense leaves of Rhizophora apiculata produce a large amount of litter compared to species with small and narrow leaves (Efriveldi et al. 2021). Leaf litterers deposited on the mangrove soil mainly consisted of ungrazed material largely processed through a detritus-based food chain and accounts for considerable near-shore secondary productivity (Odum & Heald 1975). Heavy lingo cellulose deposition is an adaptation possessed by the

leaves of Rhizophora apiculata to survive in any harsh environmental fluctuations; these depositions make the leaf hard and slow down the degradation process (Nielsen & Andersen 2003). Degradation of fallen mangrove vegetation starts immediately after its colonization by fungi and bacteria (Alongi et al. 1989; Moran & Hodson 1989). Fungal saprophytic enzymes deteriorate the lignocellulose compounds of mangrove leaves (Findlay 1986) and the degraded mangrove vegetative materials support the colonization of heterogeneous microbial communities (Odum & Heald 1975; Bano et al. 1997). Microscopic examination of vegetative matter reveals the presence of a complex community composed of fungi, bacteria, protozoa, and microalgae (Odum & Heald 1975) for their growth mangroves absorb the essential nutrients released by these organisms (Alongi et al. 2003; Reef et al. 2010; Srisunont et al. 2017). Therefore, nutrient cycling by litterfall does not only promote nutrient recycling in mangroves but also strengthen the bonds between different trophic levels in the mangrove chain (Srisunont et al. 2017).

Ciliates associated with mangrove leaf litters such as Oligotrichia, Hypotrichia and Peritricha were the important detritivores that showed a characteristic role in nutrient cycling (Utz 2008; Li et al. 2010). They are the main bacterial consumers who showed some special characteristics such as modified oral cilia, short generation times, and rapid multiplication (Li et al. 2010). Ekelund et al. (2002) experimentally showed that the organic matter released by plants could stimulate bacterial and ciliate activity in the root zone leading to mineralization of organic soil nitrogen and assimilation by plants. The activity of ciliates may be important in mangrove plant nutrition; on other hand, the growth of plants may significantly affect the ciliate community. Kathiresan & Bingham (2001) have extensively studied the role of benthic communities on the growth of the mangrove ecosystem in the Pichavaram mangrove ecosystem in India.

Major factors influencing ciliate diversity are tidal inundations, local environmental conditions and physicochemical characteristics of water (Dorothy et al. 2003). Each of these factors individually contributes to the diversity and abundance of mangrove ciliates (Chithra & Sunil Kumar 2015, 2018, 2019). Ciliates are found attached to different microhabitats of the mangrove ecosystem, but their diversity and abundance depend on the nature of substrate they attach. Other than the nature of substrate, decomposition time of the attached substrate and leaching should play a significant role in determining the initial colonizers and later the protozoan species especially ciliates Dorothy et al. (2003). Most of the studies discussing the ciliate diversity were reported from the northern regions of India (Das et al., 1993; Piyali & Das, 1997; Mahajan & Nair 1965; Das 1971; Bindu 2010). There are no previous studies examined the diversity of ciliates on the sedimented leaf litters of the southern coast of India. Thus, the objectives of this study were to investigate the species composition, diversity and density of ciliates on the sedimentary leaf litters with respect to the seasonal hydrological parameters.

Materials and Methods

Study area

The study area was located in Ayiramthengu mangrove situated (lat. 9° 6' to 9° 8' N long. 76° 2' to 76° 29' E) in Kollam district of Kerala in the part of the Kayamkulam estuary, which is the narrow stretch of tropical backwater on the southwest coast of India (Figure 1a). The area covers about 20 acres, and most of the inner regions of mangrove forest patches were left untouched by humans. Two streams and a canal enter the lake during the rainy season, which links the rivers Pamba and Achankoil. Ten sampling sites were considered for the study, and each site was fixed at an interval of 50 m.

Hydrology

The water samples were collected in a properly labelled clear airtight container. The temperature was measured on-station using portable mercury thermometer (-20°-110° C). Water samples were transferred to the lab in dark conditions, maintaining at 4°C for the determination of other physico chemical parameters. pH of the sample was analyzed using a pH meter (MK 6) and a conductivity meter (EQ 660A) was used to analyse the presence of electrolytes in the sample. The Winkler method was used to determine the amount of dissolved oxygen in the sample. Mohr's method was used to analyze the presence of chloride and the Phenol disulphonic acid method was used to analyze the presence of nitrate concentration in mangrove water. The stannous chloride method was used to analyze the presence of phosphate and turbidimetric method was used to analyze the presence of sulphate concentration in mangrove water. All the above-mentioned analysis was carried out according to the standard method prescribed in American Public Health Association et al. (1998).

Collection, identification and enumeration of epibenthic ciliates

The methods described by Maybruck & Rogerson (2004) were adopted for the collection of epibenthic organisms from mangrove leaf litter. Mangrove leaf litters were covered with a heterogeneous mixture of trapped material (Figure 1b). Samples were collected by taking one cm^2 patch of the moist film of trapped

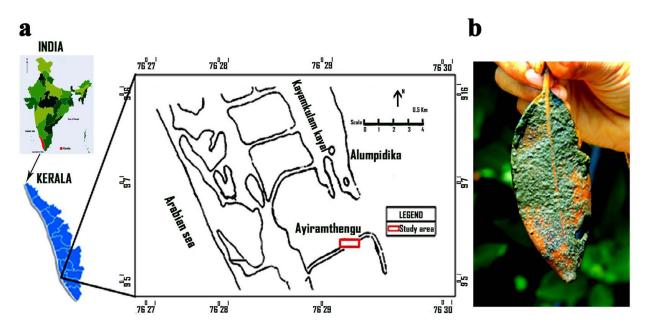


Figure 1. Map of Ayiramthengu mangrove ecosystem (a); Mangrove leaf litter with heterogeneous trapped material (b)

material to a sterile 15 ml sampling bottle. Continuous one-year sampling (June 2014 - May 2015) was carried out to analyze the composition and distribution of epibenthic organisms in mangrove leaf litter. Biometrical characteristics of the epibenthic organisms collected from the mangrove leaf litter were carefully examined under live condition. For detailed analysis, samples were treated with silver carbonate technique proposed by Fernandez-Galiano (1976). The data collected were analyzed using PAST 4.03 (Hammer *et al.* 2001), statistical software packages for Social Sciences (SPSS) software version 16.0 (Handcock *et al.* 2008) and Biodiversity pro (McAleece *et al.* 1997).

Results

Environmental factors

The overview of hydrological parameters analysed during the study period is provided in Tab. 1. Temperature and dissolved oxygen record their minimum values in post-monsoon. Temperature exhibits its maximum values in pre-monsoon, and dissolved oxygen records its maximum values in monsoon. Conductivity, pH and salinity exhibit their minimum values in the monsoon and maximum values in the pre-monsoon. Nitrate, phosphate and sulphate exhibit their minimum values in pre-monsoon and maximum values in monsoon. The average annual precipitations relative to the baseline time periods of June 2014 to May 2015 are presented in Table 1. The data reveals that the maximum average rainfall (374.97 mm) was recorded in the monsoon and the minimum (100.47 mm) recorded post-monsoon.

Species composition, dominance index and density

An aggregate of 115 species spread over 16 subclasses was identified during the one-year study period. Among them, Peritricha represents the subclass having the maximum species number (64) followed by Suctoria (11). Heterotrichea, Hypotrichia, Nassophorea and Stichotrichia are the subclasses having only one species record during the entire study period. Vorticella companula Ehrenberg, 1831 records the maximum abundance (1480 no./cm²) and dominance index (0.0460) followed by Euplotes minuta (1349 no./cm² & 0.04198). Epiphyllum soliforme, Acanthocustis aculeate, Cothurnia recurvate, Pyxicola annulata, Vorticella campanulate, Cohnilembus sp.1, Tintinnopsis sacculus, Acineta calkinsi and Ephelota plana are with the rarest distributional records from Aviramthengu mangroves. Cothurnia represents the genus with a maximum number of species (18) followed by Zoothamnium (11) (Table 2).

Out of 32,132 individuals counted during the one year survey, maximum numbers of ciliates were recorded during post-monsoon (14431) trailed by monsoon (10403) and pre-monsoon (7298). Peritricha represents the subclass having the maximum number of individuals (19778) followed by Suctoria (4026) and Spirotrichea (3143) (Table 3). Percentage distribution pattern also highlights that 62% of the total ciliate composition in the Ayiramthengu mangrove ecosystem is occupied with Peritricha followed by Suctoria (13%) and Spirotrichea (10%) (Figure 2). Among the recorded peritrichs, 45% of them are distributed in post-monsoon, 32% in monsoon and 23% in pre-monsoon (Figure 2a). Investigating the species dispersion pattern, 93 species

	Monsoon (M)	Post- monsoon (P.M)	Pre-monsoon (Pr. M)	Minimum	Maximum	
Temperature	28.99 ± 0.32	27.85 ± 0.92	29.05 ± 1.36	26.5°C (P.M)	32.7°C (Pr. M)	
pН	6.85 ± 0.43	7.14 ± 0.09	7.53 ± 0.09	6.2 (M)	8.2 (Pr. M)	
Conductivity	12.94 ± 8.62	26.64 ± 10.37	27.45 ± 11.91	0.2 s/m (M)	37.4 s/m (Pr. M)	
D.O	4.62 ± 0.39	3.80 ± 0.85	4.59 ± 1.03	0.8 mg/l (P.M)	7.6 mg/l (M)	
Salinity	12.15 ± 1.28	14.00 ± 1.04	17.56 ± 2.86	10 ppm (M)	25.2 ppm (Pr. M)	
Nitrate	2.47 ± 0.65	2.32 ± 0.59	1.64 ± 0.98	0.09 mg/l (Pr. M)	7.74 mg/l (M)	
Phosphate	2.55 ± 0.05	2.49 ± 0.06	2.33 ± 0.02	2.26 mg/l (Pr. M)	2.87 mg/l (M)	
Sulphate	267.36 ± 41.02	126.41 ± 59.17	104.10 ± 77.20	7.94 mg/l (Pr. M)	295.18 mg/l (M)	
Average Rainy Days	17	7	6			
Average Rain Fall (mm)	374.97	100.47	183.4			

Table 1.	Overview	of hydro	logical	parameters

were recorded during the monsoon, 111 during the postmonsoon and 105 during the pre-monsoon.

Diversity indices

The maximum Shannon and Simpson diversity was observed in the monsoon (0.637, 0.398) and the minimum recorded during the pre-monsoon (0.588, 0.432). Analyzes of species richness by Margaleff index, showed maximum species richness in pre-monsoon (4.142) and the lowest recorded in post-monsoon (3.847). Maximum value of Shannon Evenness index was observed in monsoon (0.529) and minimum was recorded during pre-monsoon (0.488) (Table 4).

Analyzing the subclass wise diversity index, Suctoria (1.017), Stichotrichia (1.016) and Peritricha (1.015) were with the maximum Shannon diversity and Heterotrichea (0.843) and Cyrtophoria (0.853) with the minimum diversity. Maximum Simpson diversity was observed in the case of Stichotrichia (0.091) and the minimum diversity was observed in the case of Cyrtophoria (0.176). Similar to the Shannon diversity index, Shannon evenness also shows a similar trend,

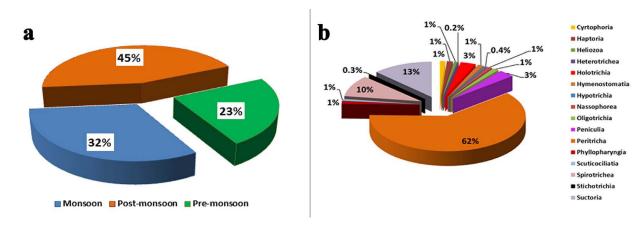
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SI. No.	CILIATES	Abundance	Dominance index
	Cyrtophoria (3)		
1	Chilodonella cucullulus	59	0.001836176
2	Chlamydodon sp1	106	0.003298892
3	Dysteria proraefrons	196	0.006099838
	Haptoria (5)		
4	Acineria incurvata	80	0.00248973
5	Epiphyllum shenzhenense	127	0.003952446
6	Epiphyllum soliforme	49	0.00152496
7	Litonotus paracygnus	88	0.002738703
8	Litonotus petzi	50	0.001556081
	Heliozoa (3)		
9	Acanthocystis aculeata	3	9.33649E-05
10	Actinophrys eichhorni	97	0.003018797
11	Actinophrys sol	108	0.003361135
	Heterotrichea (1)		
12	Blepharisma lateritium	73	0.002271879
	Holotrichia (3)		
13	Coleps hirtus	689	0.021442798

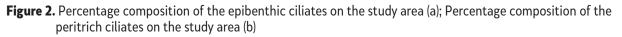
Table 2. Abundance and dominance index of ciliates counted during the study period

OL M		41 1	D ' ' I
SI. No.	CILIATES	Abundance	Dominance index
14	Holophyra oblonga	92	0.002863189
15	Prorodon Sp1	246	0.007655919
40	Hymenostomatia (2)		
16	Uronea acutum	247	0.007687041
17	Uronea caudata	160	0.00497946
	Hypotrichia (1)		
18	Diophrys hystrix	141	0.004388149
	Nassophorea (1)		
19	Nassula Ornata	347	0.010799203
	Oligotrichia (2)		
20	Strobidinopsis elongata	76	0.002365243
21	Strobidinopsis paracalkinsi	341	0.010612474
	Peniculia (2)		
22	Frontonia subtropica	354	0.011017055
23	Frontonia tchibisovae	149	0.004637122
24	Paramecium brusaria	489	0.015218474
25	Paramecium polycaryum	80	0.00248973
	Peritricha (64)		
26	Cothurnia acuta	79	0.002458608
27	Cothurnia angusta	141	0.004388149
28	Cothurnia annulata	266	0.008278352
29	Cothurnia anomala	212	0.006597784
30	Cothurnia asymmetrica	582	0.018112785
31	Cothurnia carinogammari	412	0.012822109
32	Cothurnia coarctata	354	0.011017055
33	Cothurnia elegans	168	0.005228433
34	Cothurnia kahli	261	0.008122744
35	Cothurnia kiwi	267	0.008309473
36	Cothurnia limnoriae	183	0.005695257
37	Cothurnia recurva	355	0.011048176
38	Cothurnia recurvata	31	0.00096477
39	Cothurnia Sp 1	121	0.003765716
40	Cothurnia Sp 2	87	0.002707581
41	Cothurnia trophonicola	171	0.005321798
42	Cothurnia vaga	350	0.010892568
43	Cothurnia variabilis	182	0.005664135
44	Epistylis coronata	329	0.010239014
45	Epistylis hentscheli	95	0.002956554
46	Epistylis nayagarae	365	0.011359393
47	Epistylis plicatilis	267	0.008309473
48	Epistylis procumbens	245	0.007624798
49	Paravorticella sp	457	0.014222582
50	Platycola mollis	104	0.003236649

SI. No.	CILIATES	Abundance	Dominance index
51	Platycola truncata	91	0.002832068
52	Pseudovorticella banatica	364	0.011328271
53	Pseudovorticella clampi	444	0.013818001
54	Pseudovorticella stilleri	553	0.017210258
55	Pyxicola annulata	44	0.001369351
56	Pyxicola constricta	139	0.004325906
57	Pyxicola eforiana	86	0.00267646
58	Pyxicola ligae	52	0.001618324
59	Pyxicola limbata	115	0.003578987
60	Thuricola folliculata	107	0.003330014
61	Thuricola gracilis	193	0.006006473
62	Thuricola incisa	83	0.002583095
63	Thuricola valvata	84	0.002614216
64	Vaginicola ceratophylli	63	0.001960662
65	Vaginicola compressa	84	0.002614216
66	Vaginicola elongata	112	0.003485622
67	Vaginicola gigantea	76	0.002365243
68	Vaginicola inclinata	51	0.001587203
69	Vaginicola ingentia	64	0.001991784
70	Vaginicola vestita	120	0.003734595
71	Vorticella alba	1181	0.036754637
72	Vorticella annulata	564	0.017552596
73	Vorticella bidulphae	733	0.02281215
74	Vorticella campanulate	48	0.001493838
75	Vorticella companula	1480	0.046060002
76	Vorticella longitricha	525	0.016338852
77	Zoothamnium duplicatum	867	0.026982447
78	Zoothamnium foissneri	429	0.013351176
79	Zoothamnium hentscheli	436	0.013569028
80	Zoothamnium kentii	788	0.024523839
81	Zoothamnium mengi	418	0.013008839
82	Zoothamnium mucedo	242	0.007531433
83	Zoothamnium parahentscheli	450	0.01400473
84	Zoothamnium pararbuscula	730	0.022718785
85	Zoothamnium plumula	699	0.021754015
86	Zoothamnium procerius	188	0.005850865
87	Zoothamnium ramosissimum	161	0.005010581
88	Zoothamnium sinense	658	0.020478028
89	Zoothamnopsis liui	172	0.005352919
	Phyllopharyngia (3)		
90	Aegyriana oliva	152	0.004730487
91	Chlamydodon mnemosine	167	0.005197311
92	Chlamydodon pedarius	56	0.001742811

SI. No.	CILIATES	Abundance	Dominance index
	Scuticociliatia (3)		
93	Cohnilembus verminus	158	0.004917216
94	Cohnilembus sp.1	44	0.001369351
95	Pleuronema wilberti	55	0.001711689
	Spirotrichea (8)		
96	Euplotes antarcticus	328	0.010207892
97	Euplotes eurystomus	107	0.003330014
98	Euplotes minuta	1349	0.04198307
99	Euplotes muscicola	111	0.0034545
100	Stentor roeseli	314	0.00977219
101	Tintinnopsis rotundata	647	0.02013569
102	Tintinnopsis sacculus	43	0.00133823
103	Tintinnopsis schotti	244	0.007593676
	Stichotrichia (1)		
104	Stichotricha secunda	106	0.003298892
	Suctoria (11)		
105	Acineta biloba	726	0.022594299
106	Acineta calkinsi	32	0.000995892
107	Acineta compressa	212	0.006597784
108	Acineta papillifera	96	0.002987676
109	Crossacineta annulata	216	0.006722271
110	Ephelota gemmipara	724	0.022532055
111	Ephelota mammillata	708	0.022034109
112	Ephelota plana	29	0.000902527
113	Ephelota turncata	438	0.013631271
114	Metacineta mystacina	71	0.002209635
115	Podophrya macrostyla	774	0.024088136
			1





Sub class	Monsoon	Post-monsoon	Premonsoon	Total
Cyrtophoria	91	246	24	361
Haptoria	115	196	83	394
Heliozoa	44	123	41	208
Heterotrichea	38	16	19	73
Holotrichia	568	230	229	1027
Hymenostomatia	182	204	21	407
Hypotrichia	66	57	18	141
Nassophorea	122	184	41	347
Oligotrichia	89	283	45	417
Peniculia	261	557	254	1072
Peritricha	6293	8847	4638	19778
Phyllopharyngia	79	126	170	375
Scuticociliatia	93	100	64	257
Spirotrichea	807	1633	703	3143
Stichotrichia	45	37	24	106
Suctoria	1510	1592	924	4026
	10403	14431	7298	32132

Table 3. Monthly distribution of different sub classes of ciliates during the study period

where also the maximum evenness was observed in the case of Suctoria (0.8446) followed by Stichotrichia (0.8438) and Peritricha (0.8429). Minimum evenness was observed in the case of Heterotrichea (0.7001). The subclass Heterotrichea showed the maximum species richness value (5.537) and the minimum observed in the case of Peritricha (2.344) (Table 4).

Relationship between ciliates and environmental factors

Analysis of variance (ANOVA) showed the relationship between different subclasses of ciliates with environmental factors. The monthly variances in the distribution of ciliates were analysed by considering p < 0.05 as significant. The observed p-value (2.7E-106) is lesser than the significance level 0.05, and showed that most of the months have different means (Table 5).

Multivariate statistical analysis

Cluster analyzes clearly showed two general clusters: monsoon and post-monsoon period as one cluster, and the other cluster refers to the community structure in other months of the year (Figure 3). Detailed analysis revealed five distinct groupings, which appeared to reflect differences in the monthly distribution of ciliates in the Ayiramthengu mangroves ecosystem (Figure 3). Cluster 1 consisted of the month

of June, July and August, which is the monsoon season, with high diversity and evenness of organisms. Cluster 2 consisted of the months of September and October, represented by the late monsoon period and early postmonsoon with unique climatic and physicochemical characteristics, with a similar population density. Cluster 3 consisted of the months of November, December and January with maximum population density and species number, Cluster 4 consisted of the months of February and March, with similar population density values as in Cluster 3. Cluster 5 consisted of the months of April and May with minimum population density, maximum richness and lowest diversity.

Canonical correspondence analysis (CCA)

CCA revealed sixteen distinct ciliate subclass groupings, which appeared to reflect differences in environmental characters within the Ayiramthengu mangroves (Figure 4). Eigen value of axis 1 and axis 2 itself explains 70.28 % of the relationship between environmental variables and ciliate communities. This result shows the close association between environmental variables and ciliate community.

In the tripod of CCA, vectors dissolved oxygen, temperature and sulphate have maximum length, and strongly influences the ciliate diversity. The ordination diagram of CCA revealed a strong negative loading of axis 1 with conductivity and salinity. Oligotrichia and Peniculia showed a negative association with axis 2 and illustrate the importance of conductivity and salinity

									sirotou2	1.01	0.84	0.10	2.80
									stichotrichia	1.01	0.84	0.09	5.10
	-								Spirotrichea	1.00	0.83	0.10	2.87
	Pre-monsoon	0.588	0.432	4.142	0.488			ß	tisilisositus2	0.97	0.81	0.11	4.23
	Pr	0.5	0.4	4.1	0.4			ßig	Phyllopharyn	1.01	0.83	0.10	3.91
									Peritricha	1.01	0.84	0.10	2.34
	u								siluɔin9¶	0.96	0.80	0.12	3.35
	Post-Monsoon	0.635	0.404	3.847	0.527				oligotrichia	0.86	0.71	0.17	3.83
	Po	0.6	0.4	3.6	0.5				Nassophorea	0.94	0.78	0.13	4.00
									нуроттісһіа	0.89	0.74	0.13	4.70
								ıatia	motson9myH	0.87	0.72	0.15	3.89
' period	Monsoon	0.637	0.398	3.983	0.529				Bidzirichia	0.98	0.81	0.11	3.38
the study	Me	0.0	0.	3.0	0.5	_		e	Heterotriche	0.84	0.70	0.14	5.53
d during .						udy period			вохоіІ9Н	0.95	0.79	0.12	4.31
s recorde						ring the st			Haptoria	1.00	0.83	0.10	3.89
ty indices						scorded du			cyrtophoria	0.85	0.70	0.17	3.94
Table 4. Sub class wise diversity indices recorded during the study period	Index	Shannon H' Log Base 10.	Simpsons Diversity (D)	Margaleff M Base 10.	Shannon Evenness	Sub class wise diversity indices recorded during the study period				Shannon H' Log Base 10.	Shannon Evenness	Simpsons Diversity (D)	Margaleff M Base 10.

Table 5. ANOVA table showing the monthly distributional variations of epibenthic ciliates in relation to hydrological parameters

Source of Variation	SS	df	MS	F	P-value	F crit
Between Groups	31112248	23	1352706	82.20345	2.7E-106	1.570294
Within Groups	4344276	264	16455.59			
Total	35456524	287				

in their abundance and distribution. The lowest level of phosphate, nitrate, salinity and conductivity favors the influx of Spirotrichea, Nassophorea, Haptoria and Peritricha in mangrove leaf litter. Holotrichia reaches its maximum abundance in higher sulphate concentration and Stichotrichia reaches its maximum number in a moderate temperature range. Similarly, Phyllopharyngea reaches its maximum abundance in mid-low pH and Peniculia reaches its maximum number in moderate salinity condition. Oligotrichia reaches its maximum number in mid-low conductivity.

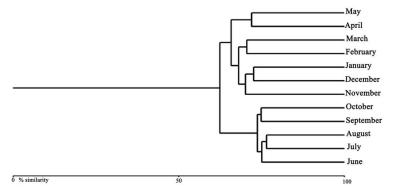
k-dominance

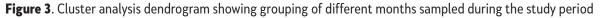
A k-dominance plot curve was drawn based on the community structure of ciliates in the Ayiramthengu mangrove ecosystem. In the present research, the data collected during the one-year survey was fed into the dominance plot (Figure 5). The highest dominance was seen in the month of May, where the ciliate assemblage had the highest diversity.

Discussion

The present investigation describes species composition, distribution, diversity and abundance of epibenthic ciliates from the sedimentary leaf litters of Ayiramthengu mangroves in relation to the abiotic factor prevailing in the ecosystem. Several studies have been conducted on the physicochemical analysis of mangrove water from various parts of India, and only a few studies have been reported on the taxonomy and community structure of ciliates from Indian mangroves (Dorothy et al. 2003; Nandi et al. 1993; Sarkar 2014; Chithra & Sunil Kumar 2015, 2018, 2019a, 2019b; Chithra et al. 2018). Monthly variations in the distribution of epibenthic ciliates in mangrove leaf litters were well documented by analyzing the level of significance through one-way ANOVA and CCA. The analysis of the data revealed that the epibenthic ciliates attach to the mangrove soil exhibit monthly variations according to the changes in hydrological conditions. Chen et al. (2009) had made a similar study in the tropical mangrove ecosystem and recorded that the changes in hydrological parameters determine the distribution of mangrove species in each habitat.

Mangrove soils are typically saline, anoxic, acidic, and frequently waterlogged. These soil properties directly affect the distribution of mangrove ciliates (Li *et al.* 2010). In the present study, conductivity and dissolved oxygen exhibited high negative correlations with ciliates. The conductivity and the dissolved oxygen frequently affect ciliates distribution (Ekelund & Ronn 1994; Opravilová & Hájek 2006; Ehrmann *et al.* 2012) and it affects ciliate species composition, as well as ciliate species diversity and density in the soil (Lara *et al.* 2016). Very few studies have been reported to investigate the ciliate community structure and physicochemical properties (Chao *et al.* 2006; Aguilera *et al.* 2006; Li *et al.* 2010; Ting *et al.* 2012; Fokam *et al.* 2015; Debastiani *et al.* 2016) and the information is still lacking from India.





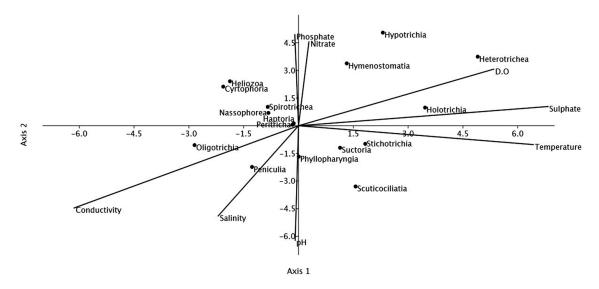


Figure 4. CCA plot similarity showing grouping of sub classes sampled during different seasons along with environmental factors

Peritrichs are the dominant ciliate subclass recorded from the study area. They are adapted to a wide range of habitats. Sedentary forms are abundant in marine, brackish and freshwater habitats attaching to a wide variety of substrates, plant, animals and inanimate (Warren 1983). Most of the peritrichs recorded from the study are attached to algal communities, mostly on Spirogyra. Interspecific factors such as growth pattern and cyst formation may also be highly significant in the case of peritrichs. Pseudo colony formation and solitary life in peritrichs may provide survival value by enhancing feeding efficiency, surface dominance, conjugation and predator avoidance (Spoon 1974).

Suctorians are the second dominant epibionts observed from the study habitat, similar to peritrichs, most of the suctorians are attached to a living or nonliving substratum. Suctorians are very common both in marine and freshwater environments (Foissner *et al.* 2007). They attach to their hosts directly or by using stalks, and they feed mainly on other ciliates (Verni & Guatieri 1997). While analyzing the species-wise abundance, Vorticella companula Ehrenberg, 1831 shows the maximum abundance and dominance index. During unfavourable conditions, they develop a posterior ring of cilia to become a telotroch. It separates away from the stalk and swims away to some favourable place where it develops a stalk and starts a normal life. The telotroch helps in its dispersal. Vorticella companula Ehrenberg, 1831 is a solitary species and typically form pseudocolonies because of telotrochs settling on the same substrate and very close to the mature organisms (Paranjape 1987). Euplotes minuta is the second most dominant species found to be dominant in the Ayiramthengu mangrove ecosystem. It is a highly diversified and cosmopolitan genus, with a large number of species that have been observed and investigated in all kinds of biotopes (Chen et al. 2013). Zoothamnium can be found in freshwater, brackish and marine waters between 0 - 8 meters deep and typically form a symbiotic relationship with a wide variety of animals, although some may be free-living and attaching to aquatic plants and inanimate substrates (Clamp et al. 2006).

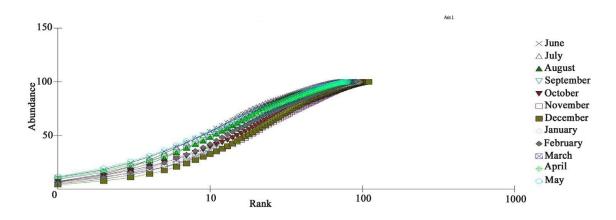


Figure 5. K dominance plot showing the abundance of ciliates in study year

Analyzing the species composition of ciliates with respect to three seasons, post-monsoon represents the season with maximum species number followed by pre-monsoon. Thus, the dominant occurrence of the above-mentioned ciliates in each season may be closely associated with the species-specific environmental conditions that are required for encystment or excystment (Godhantaraman 2002). Especially in biological and other characteristic features of these ciliates support their dominance in the dynamic mangrove habitat. The present study reports the occurrence of 31 species of ciliates which were evenly distributed throughout the study period. It is because of their ability to switch to a different diet throughout the three seasons (Pierce & Turner 1992). During the study period, maximum Evenness, Shannon and Simpson diversity values were observed in monsoon and minimum recorded in pre-monsoon, due to the impact of the highest species richness in pre-monsoon. Similar studies were reported from the Vellar mangrove waters, where the total species richness of ciliates was highest in pre-monsoon and lowest during monsoon, as commonly observed in many marine coastal and estuarine waters (Capriulo & Carpenter 1983; Verity & Lagdon 1984; Verity 1987; Paranjape 1987; Burkill et al. 1995; Kamiyama & Tsujino 1996). Similar seasonal variations have also been reported in the Parangipettai coastal waters (Krishnamurthy & Santhanam 1975; Naidu et al. 1977; Mahajan & Nair 1965). Cluster analysis and k-dominance index also revealed the similar diversity, abundance and distribution pattern of ciliates during one-year study period. Studies on ANOVA records a station-wise heterogeneity during the oneyear study period. Analysing the species diversity and evenness, Suctoria, Stichotrichia and Peritricha showed maximum diversity values, while Heterotrichea and Cyrtophoria the minimum diversity. Suctorians can be found in all types of water bodies on a wide diversity of hosts and substrates. Larger ciliate subclasses such as Heterotrichea and Cyrtophoria were found occasionally in the sampling area because the species of this subclass are more adapted to freshwater habitat (Mahajan & Nair 1965). The majority of these ciliates are commensals of various water invertebrates or vertebrates (Dovgal 2002). Taxonomic studies highlighting the distributional records of the above-mentioned groups were reported earlier from the study habitat (Chithra & Sunil Kumar 2015, 2018, 2019a, 2019b; Chithra et al. 2018). When analyzing the CCA, vectors for strong positively and negatively associated variable showed an acute angle and were close to each other. Values of positive and negative loading axis 1 and 2 of CCA helps to understand the effect of environmental variables to the distribution of mangrove ciliates. Analysing the ordination plot for CCA, vectors such as conductivity, temperature, sulphate and dissolved oxygen showed maximum length that, strongly influence ciliate composition and abundance. Habitat preference of ciliate was according to the changes in suspended materials and the changes favors the occurrence of bacterivores, algivorous and omnivorous ciliates in the study habitat (Foissner & Berger 1996).

Conclusion

The present study highlighted the role of physicochemical parameters in structuring the ciliate community in the Ayiramthengu mangrove ecosystem. Ciliates, generally considered among the most evolved and complex protozoa, which can perceive and react to a wide variety of environmental stimuli. The study also revealed that the ecological variations recorded from the mangrove habitat were very significant to make a change in the ciliate distribution. Meanwhile, in an aspect of ciliate as ecological indicators, evenly distributed community structure reflects the existence of a healthy food web in the present ecosystem. The literature survey revealed the fact that little work has been done on the diversity and distribution of ciliates on mangrove leaf litters of Rhizophora apiculata, and our current finding may provide some information for researchers in the field of mangrove ciliates.

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Declarations

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References

- Aguilera, A., Manrubia, S.C., Gómez, F., Rodríguez, N., Amils, R. (2006). Eukaryotic community distribution and its relationship to water physicochemical parameters in an extreme acidic environment, Río Tinto (Southwestern Spain). *Applied and Environmental Microbiology*, **72** : 5325-5330.
- Alongi, D. M., Boto, K. G., Tirendi, F. (1989). Effect of exported mangrove litter on bacterial productivity and dissolved organic carbon fluxes in adjacent tropical nearshore sediments. *Marine ecology progress series*. *Oldendorf*, 56(1): 133-144.

- Alongi, D. M., Clough, B. F., Dixon, P., Tirendi, F. (2003). Nutrient partitioning and storage in arid-zone forests of the mangroves *Rhizophora stylosa* and *Avicennia marina*. *Trees*, **17**(1): 51-60.
- American Public Health Association, American Water Works Association, Water Pollution Control Federation, & Water Environment Federation. (1998). Standard methods for the examination of water and wastewater (Vol. 2). American Public Health Association.
- Bano, N., Nisa, M. U., Khan, N., Saleem, M., Harrison, P. J., Ahmed, S. I., Azam, F. (1997). Significance of bacteria in the flux of organic matter in the tidal creeks of the mangrove ecosystem of the Indus River delta, Pakistan. *Marine ecology progress series*, **157**: 1-12.
- Bindu, L. (2010). Freshwater Ciliates (Protozoa) from Kolkata Wetlands. *Records of the Zoological Survey of India*, **110**(2), 81-88.
- Burkill, P. H., Edwards, E. S., Sleight, M. A. (1995). Micro zooplankton and their role in controlling phytoplankton growth in the marginal ice zone of the Bellingshausen Sea. *Deep Sea Research Part II: Topical Studies in Oceanography*, **42**(4-5): 1277-1290.
- Capriulo, G. M., Carpenter, E. J. (1983). Abundance, species composition and feeding impact of tintinnid microzooplankton in central Long Island Sound. *Marine* ecology progress series. Oldendorf, **10**(3): 277-288.
- Chao, A., C. Li, P., Agatha, S., Foissner, W. (2006). A statistical approach to estimate soil ciliate diversity and distribution based on data from five continents. *Oikos*, **114**(3): 479-493.
- Chen, X., Zhao, Y., Al-Farraj, S. A., Al-Quraishy, S., El-Serehy, H. A., Shao, C., Al-Rasheid, K. A. (2013). Taxonomic descriptions of two marine ciliates, Euplotes dammamensis n. sp. and Euplotes balteatus (Dujardin, 1841) Kahl, 1932 (Ciliophora, Spirotrichea, Euplotida), collected from the Arabian Gulf, Saudi Arabia. Acta Protozoologica, 52(2).
- Chen, Q. H., Tam, N. F. Y., Shin, P. K., Cheung, S. G., Xu, R. L. (2009). Ciliate communities in a constructed mangrove wetland for wastewater treatment. *Marine pollution bulletin*, 58(5):711-719.
- Chithra, P., Sunil Kumar, R. (2015). Preliminary studies on epibiotic protista in the mangrove ecosystem of Ayiramthengu, Kerala coast. *Journal of the Marine Biological Association of India*, **57**(2): 90-94.
- Chithra, P., Sunil Kumar, R., Nisanth, R. (2018). First report of peritrich ciliates of genera Pseudovorticella from the Ayiramthengu mangrove ecosystem, southwest coast of India. International Journal for Research in Applied Science and Engineering Technology, 6(3): 2888-2892.
- Chithra, P., Sunil Kumar, R. (2019a) Abstract Proceedings, ICBN. Kochi, Distribution and diversity of ciliate epibionts on the prop root of Rhizophora apiculata, pp.164.
- Chithra P., Sunil Kumar, R. (2019b) New distributional records of eighteen species of Vorticella (Ciliophora:

Peritricha) from mangrove ecosystem of Ayiramthengu in south west coast of India. *Journal of the Marine Biological Association of India*, **61**: 100-105.

- Chithra, P., Sunil Kumar, R., Nisanth, R. (2018). First report of peritrich ciliates of genera Pseudovorticella from the Ayiramthengu mangrove ecosystem, southwest coast of India. International Journal for Research in Applied Science and Engineering Technology, 6(3): 2888-2892.
- Clamp, J. C. (2006). Redescription of Lagenophrys cochinensis Santhakumari Gopalan, 1980 (Ciliophora, Peritrichia, Lagenophryidae), an ectosymbiont of marine isopods, including new information on morphology, geographic distribution, and intraspecific variation. *Journal of Eukaryotic Microbiology*, **53**(1): 58-64.
- Das, A.K. (1971). Leptopharynx chlorophagus sp. nov. (Ciliata: Protozoa) from freshwater of West Bengal, India. *Current Science.*, **40** : 195-196.
- Das, A.K., MandaI, A.K., Sarkar, N.C. (1993). Free living Protozoa. Zool. Surv. India, Fauna of West Bengal, State Fauna Series, **3**: 1-133.
- Debastiani, C., Meira, B. R., Lansac-Tôha, F. M., Velho, L. F. M., Lansac-Tôha, F. A. (2016). Protozoa ciliates community structure in urban streams and their environmental use as indicators. *Brazilian Journal* of Biology, 76: 1043-1053.
- Dorothy, K. P., Satyanarayana, B., Kalavati, C., Raman, A. V. (2003). Protozoa associated with leaf litter degradation in Coringa mangrove forest, Kakinada Bay, east coast of India. *Indian Journal of Geo-Marine Sciences* 32: 45-51.
- Dovgal, I. V. (2002). Evolution, phylogeny and classification of Suctorea (Ciliophora). *Protistology*, **2** (4): 194-270.
- Duke, N. C. (1993). Mangrove floristics and biogeography. *Coastal and Estuarine studies*, 63-63.
- Ehrmann, O., Puppe, D., Wanner, M., Kaczorek, D., Sommer, M. (2012). Testate amoebae in 31 mature forest ecosystems-densities and micro-distribution in soils. *European journal of protistology*, **48**(3): 161-168.
- Ekelund, F., Ronn, R. (1994). Notes on protozoa in agricultural soil with emphasis on heterotrophic flagellates and naked amoebae and their ecology. *FEMS microbiology reviews*, **15**(4), 321-353.
- Ekelund, F., Frederiksen, H. B., Rønn, R. (2002). Population dynamics of active and total ciliate populations in arable soil amended with wheat. *Applied and environmental microbiology*, **68**(3): 1096-1101.
- Efriyeldi, E., Amin, B., Hersa, T. (2021). Production of Rhizophora Mangrove Leaf Litter in The Sungai Bersejarah Mangrove Ecosystem, Siak Regency. In *IOP Conference Series: Earth and Environmental Science* (Vol. 934, No. 1, p. 012073). IOP Publishing.
- Fernandez-Galiano, D. (1976). Silver impregnation of ciliated protozoa: procedure yielding good results with the pyridinated silver carbonate

method. *Transactions of the American Microscopical Society*, 557-560.

- Findlay, R. H. (1986). Biochemical indications of the role of fungi and thraustochytrids in mangrove detrial systems. *The biology of marine fungi*, 91-103.
- Foissner, W., Berger, H. (1996). A user-friendly guide to the ciliates (Protozoa, Ciliophora) commonly used by hydro biologists as bio indicators in rivers, lakes, and waste waters, with notes on their ecology. *Freshwater biology*, **35**(2): 375-482.
- Foissner, W., Chao, A., Katz, L. A. (2007). Diversity and geographic distribution of ciliates (Protista: Ciliophora). In *Protist diversity and geographical distribution* (pp. 111-129). Springer, Dordrecht: 111-129.
- Fokam, Z., Nana, P. A., Ngassam, P., Bricheux, G., Vigues, B., Bouchard, P., Sime-Ngando, T. (2015). Soil physicochemical parameters affecting abundance and distribution of Dicoelophrya nkoldaensis (Ciliophora: Radiophryidae) living in the gut of earthworms (Annelida: glossoscolecidae) collected in Bambui (nord-west Cameroon). International Journal of Current Research, 7(6):17164-17173.
- Godhantaraman, N. (2002). Seasonal variations in species composition, abundance, biomass and estimated production rates of tintinnids at tropical estuarine and mangrove waters, Parangipettai, southeast coast of India. *Journal of Marine Systems*, **36**(3-4): 161-171.
- Hammer, O., Harper, D. A., Ryan, P. D. (2001). PAST: Paleontological statistics software package for education and data analysis. *Palaeontologia electronica*, 4(1): 9.
- Handcock, M. S., Hunter, D. R., Butts, C. T., Goodreau, S. M., Morris, M. (2008). statnet: Software tools for the representation, visualization, analysis and simulation of network data. *Journal of statistical software*, 24(1): 1548.
- Hogarth, P. J. (1999). The Biology of Mangroves. J. Hogarth, P. J. (Ed). Oxford, New York: Oxford University Press. 208.
- Kamiyama, T., Tsujino, M. (1996). Seasonal variation in the species composition of tintinnid cilates in Hiroshima Bay, the Seto Inland Sea of Japan. *Journal of Plankton Research*, **18**(12): 2313-2327.
- Kathiresan, K., Bingham, B. L. (2001). Biology of mangroves and mangrove ecosystems. Advances in Marine Biology, 40: 81-251.
- Krishnamurthy, K., Santhanam, R. (1975). Ecology of tintinnids (Protozoa: Ciliata) in Porto Novo region, *Indian Journal of Geo-Marine Sciences*, **4** : 181-184.
- Lara, E., Roussel-Delif, L., Fournier, B., Wilkinson, D. M., Mitchell, E. A. (2016). Soil microorganisms behave like macroscopic organisms: patterns in the global distribution of soil euglyphid testate amoebae. *Journal of biogeography*, **43**(3): 520-532.
- Li, J., Liao, Q., Li, M., Zhang, J., Tam, N. F., Xu, R. (2010). Community structure and biodiversity of

soil ciliates at Dongzhaigang Mangrove Forest in Hainan Island, China. *Applied and Environmental Soil Science*.

- Mahajan, K. K., Nair, K. N. (1965). On some freshwater ciliates (Protozoa) from Calcutta and its environs. *Records of the Zoological Survey of India*, 63(1-4): 1-22.
- Maybruck, B.T., Rogerson, A. (2004). Protozoan epibionts on the prop roots of the Red Mangrove Tree, *Rhizophora mangle. Protistology*, **3**: 265-272.
- McAleece, N., Lambshead, P. J. D., Paterson, G. L. J., Gage, J. G. (1997). Biodiversity professional. Beta-Version. London, The Natural History Museum and the Scottish Association for Marine Sciences.
- Mini M., Lekshmy, S., Radhakrishnan, T. (2014). Kerala mangroves–Pastures of estuaries–Their present status and challenges. *International Journal of Science and Research*, **3**(11): 2804-2809.
- Moran, M. A., Hodson, R. E. (1989). Formation and bacterial utilization of dissolved organic carbon derived from detrital lignocellulose. *Limnology and* oceanography, 34(6): 1034-1047.
- Naidu, W. D., Santhanam, R., Krishnamurthy, K., Natarajan, R. (1977). The species biomass and the seasonal composition of Tintinnida (Protozoa: Ciliata). In Symposium on Warm Water Zooplankton, Goa, 1976. Proceedings. Goa, UNESCO/SCOR/NIO (pp. 520-527).
- Nandi, N. C., Das, A. K., Sarkar, N. C. (1993). Protozoa fauna of Sundarban mangrove ecosystem. *Records* of the Zoological Survey of India, **93**(1-2): 83.
- Nielsen, T., Andersen, F. O. (2003). Phosphorus dynamics during decomposition of mangrove (Rhizophora apiculata) leaves in sediments. *Journal* of *Experimental Marine Biology and Ecology*, **293**(1), 73-88.
- Odum, W. E., Heald, E. J. (1975). The detritus-based food web of an. *Estuar Res Chem Biol Estuar Syst*, **1**: 265.
- Opravilová, V., Hájek, M. (2006). The variation of testacean assemblages (Rhizopoda) along the complete baserichness gradient in fens: a case study from the Western Carpathians. *Acta protozoologica*, **45**(2): 191.
- Paranjape, M. A. (1987). Grazing by microzooplankton in the eastern Canadian Arctic in summer 1983. *Marine* ecology progress series. Oldendorf, **40**(3): 239-246.
- Pierce, R. W., Turner, J. T. (1992). Ecology of planktonic ciliates in marine food webs. *Rev. Aquat. Sci*, 6(2), 139-181.
- Piyali, C., Das, A.K. (1997). Role of protozoa in Environmental Biomonitoring, *Proceedings of the Zoological Society*. Calcutta, 50(1): 19-22
- Reef, R., Feller, I. C., Lovelock, C. E. (2010). Nutrition of mangroves. *Tree physiology*, **30**(9); 1148-1160.
- Saenger, P. (2002). Mangrove ecology, silviculture and conservation. Springer Science & Business Media. Dordrecht: Kluwer Academic Publishers: 360.

- Sarkar, S. K. (2014). Loricate ciliate tintinnids in a tropical mangrove wetland: diversity, distribution and impact of climate change. Springer.
- Spoon D (1974) Behavior of Telotrochs and Microconjugants of sessile peritrichs. J Protozool 21: 436-436.
- Srisunont, C., Jaiyen, T., Tenrung, M., Likitchaikul, M., Srisunont, T. (2017). Nutrient accumulation by litterfall in mangrove forest at Klong Khone, Thailand. *Science* & *Technology Asia*, 9-18.
- Ting, L. T., King, W. S., Hong, L. W., Ali, S. R. A. (2012). Diversity of soil protozoa (ciliates) in oil palm plantation at Sungai asap, Sarawak. In *Third International Plantation Industry Conference and Exhibition at Le Meridian Hotel, Kotakinabalu, Sabah, Malaysia.*
- Tomlinson, P. B. (1986). *The Botany of Mangroves*. Cambridge: Cambridge University Press, 413 pp.
- Utz, L. R. P. (2008). Growth of the peritrich epibiont Zoothamnium intermedium Precht, 1935 (Ciliophora, Peritrichia) estimated from laboratory experiments. *Brazilian Journal of Biology*, 68, 441-446.

- Verity, P. G. (1987). Abundance, community composition, size distribution, and production rates of tintinnids in Narragansett Bay, Rhode Island. *Estuarine, Coastal and Shelf Science*, 24(5): 671-690.
- Verity, P. G., Lagdon, C. (1984). Relationships between lorica volume, carbon, nitrogen, and ATP content of tintinnids in Narragansett Bay. *Journal of Plankton Research*, 6(5): 859-868.
- Verni, F., Gualtieri, P. (1997). Feeding behaviour in ciliated protists. *Micron*, **28**(6): 487-504.
- Vidyasagaran, K., Ranjan, M.V., Maneeshkumar, M., Praseeda, T.P. (2011). Phytosociological analysis of Mangroves at Kannur district, Kerala. *International Journal of Environmental Sciences*, **2**: 671-677.
- Vijayan, V., Rahees, N., Vidyasagaran, K. (2015). Floristic diversity and structural analysis of mangrove forests at Ayiramthengu, Kollam district, Kerala. *Journal of Plant Development Sciences* 7(2): 105-108.
- Warren, A. (1983). The ecology, morphology and taxonomy of freshwater peritrich ciliates. University of Surrey (United Kingdom).